

Changes in the ageing brain in health and disease

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Changes in the ageing brain in health and disease

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SUMMARY

The brains of individuals, who are cognitively normal, show age-related changes that include an overall reduction in brain volume and weight, which are associated with gyral atrophy and widening of the sulci of the cerebral cortex, and enlargement of the brain ventricles. These changes are partly the result of nerve cell loss but accurate estimates of neuronal loss are notoriously difficult to make. Microscopically, there are increasing amounts of the age-related pigment, lipofuscin, granulovacuolar degeneration in neurones, Hirano bodies, variable amounts of diffuse deposits of β -amyloid in the parenchyma, the presence of neurofibrillary tangles mainly confined to the hippocampus and amygdala, and sparse numbers of senile plaques in these brain regions and also in other cortical areas. Of these changes, neurofibrillary tangles and senile plaques are the neuropathological hallmark of Alzheimer's disease in which they are more abundant and widespread. Alzheimer's disease has therefore been regarded as accelerated brain ageing; however, the realization that there is a strong genetic contribution to developing the disease at least implies that it may not be the inevitable, even if frequent, consequence of old age. Understanding the molecular basis of plaque and tangle formation is advancing greatly and is the main focus of research into the cellular and molecular changes observed in the ageing brain.

1. ANATOMICAL CHANGES IN THE BRAIN ASSOCIATED WITH NON-PATHOLOGICAL AGEING

At a gross level, there is a decrease in brain volume and weight in individuals over the age of 60 years. The most obvious changes are gyral atrophy, widening of sulci, increase in ventricular volume and there is a loss of hippocampal volume (figure 1). These changes all occur in individuals who are not cognitively impaired and begin after the age of 50 years with a subsequent progressive 2-3% loss in brain weight per decade. The loss of brain substance with age is both in grey and white matter and, for example, magnetic resonance imaging studies (MRI) have demonstrated that the ratio of grey to white matter varies with age, there being relatively greater loss of white matter over the age of 50 (Harris et al. 1994). This observation is interpreted as indicating loss of myelinated axons, the axons having a greater volume than the cell body and dendrites of the neurones that are lost, the latter two neuronal compartments being located in the grey matter (Esiri et al. 1997).

There have been many studies of brain ageing in which neuronal numbers have been estimated in different brain regions. Alterations in normal ageing are still controversial and compounded by a variety of technical problems, as well as shrinkage of neuronal cell bodies such that they may not be counted within a particular class of neurones but, nevertheless, are still present (reviewed in Esiri *et al.* (1997)). However, there is a consensus that neurones in certain brain regions are lost with age (those in the hippocampus, cerebral cortex, amygdala being amongst the regions affected), whereas, other neurones in the cerebral cortex show shrinkage without loss and yet other brain regions seem to be spared of shrinkage and neuronal loss (including the nucleus basalis, an area that becomes involved in Alzheimer's disease in which the neurones do show signs of shrinkage and loss) (Chiu et al. 1984). Estimates of the degree of neuronal loss vary between reports, but are in the range of 6-25% neuronal loss in the neocortex up to the age of 100 years (Terry et al. 1987; Hansen et al. 1988). Neuronal loss in the hippocampus seems to be greater, being up to about 50% over the age range of early teens to 85 years (West 1993).

Changes in dendrites and synapses are other putative mechanisms that impair neuronal function. The evidence suggests, however, that there is dendritic growth in the normal ageing brain and that this may be a compensatory mechanism for neuronal loss (Coleman & Flood 1987). On the other hand, loss of synapses (up to 20%) seems to occur in some cortical regions (Masliah *et al.* 1993), but there is evidence that synaptic density in the hippocampus is not reduced with normal ageing in areas that receive input from layer II neurones of the entorhinal cortex where neuronal loss occurs (Lippa *et al.* 1992). This was suggested to be the consequence of sprouting by the remaining neurones, again as a compensatory reaction to neuronal loss.

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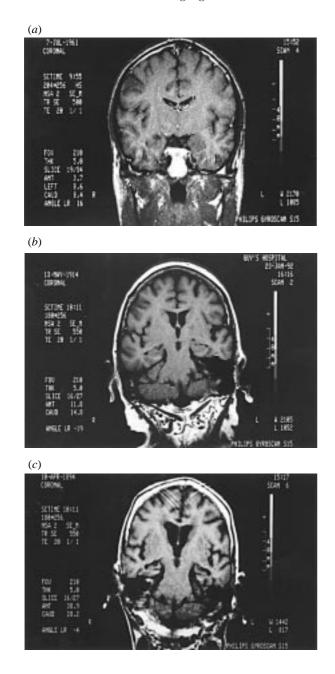


Figure 1. Magnetic resonance images of healthy and Alzheimer's disease human brains. Coronal MRI scans of the brains of (a) a healthy 35-year-old male; (b) a healthy 82-year-old female; (c) a 99-year-old female with Alzheimer's disease.

2. CELLULAR CHANGES (a) Accumulation of pigment

Lipofuscin is a pigment that accumulates in some neurones with increasing age. Lipofuscin contains peroxidized protein and lipids, and may represent increasing failure of cells to eliminate these products of peroxidation-induced cell damage. It is not clear if lipofuscin is detrimental to cells since it accumulates in the inferior olive in infancy and there is apparently no loss of these neurones throughout life (Monagle & Brody 1974). Neuromelanin, a related pigment, does appear to be associated with neuronal loss, particularly in the substantia nigra (Mann & Yates 1979).

(b) Neurofibrillary tangles

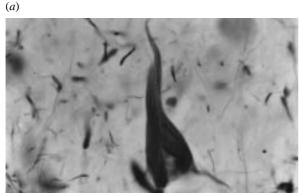
Neurofibrillary tangles and senile plaques (see below) are regarded as the histopathological hallmarks of Alzheimer's disease (figure 2). However, they are both present in the brains of non-demented individuals as, indeed, are Hirano bodies and granulovacuolar degenerative changes, the distinguishing feature of Alzheimer's disease being that the numbers of all four features are greater and more widespread. In normal ageing, the numbers of tangles, which occupy the cell body of affected neurones, is therefore relatively low and restricted to the hippocampus, amygdala and entorhinal cortex. However, in a series of brains from demented and non-demented individuals in middle to late life, it has been shown that when minimal neurofibrillary tangles are present, they are found in the transentorhinal region and in brains with increasing neurofibrillary change, the entorhinal cortex is also affected and further pathology includes the hippocampus, with associated cognitive impairment (Braak & Braak 1991). Severely demented individuals are found to have widespread cortical neurofibrillary changes. These observation have led to the proposal that neurofibrillary changes can be staged, with the early stages (I and II) not being associated with dementia.

Ultrastructurally, neurofibrillary tangles composed of paired helical filaments (PHF) and occasional straight filaments, and studies with biopsy specimens from Alzheimer's patients has shown that in neurones severely affected by PHF, the normal cytoskeleton of microtubules and neurofilaments is totally lost (Flament-Durand & Couck 1979; Gray et al. 1987). It is inconceivable that a neurone, or indeed any cell, could survive without a functional cytoskeleton and the presence of ghost or tombstone tangles in postmortem brain specimens with Alzheimer's disease, testifies to neuronal loss. Thus, some neuronal loss is associated with the presence of neurofibrillary tangles in Alzheimer's disease but this may not be a major cause of neuronal loss in normal ageing.

(c) Senile plaques

Senile plaques are areas of grey matter up to 200 µm across consisting of a central extracellular core of amyloid surrounded by swollen abnormal neurites; these plaques are also known as neuritic plaques (figure 2). The central core contains numerous proteins but the principal protein is a small peptide, 39-43 amino acids long, known as β -amyloid or A β , that is aggregated into fibrils. Small numbers of neuritic plaques are present in the normal aged brain and in Alzheimer's disease their numbers are greatly increased. However, large amounts of $A\beta$ are found as 'diffuse deposits' without a neuritic involvement in some brains from individuals who were intellectually intact, although some studies claim that there is a correlation between cognitive decline and total $A\beta$ load in the brain (reviewed in Dickson (1996)). The abnormal neurites surrounding the $A\beta$ core contain PHF and lack normal cytoskeletal numerous structures.

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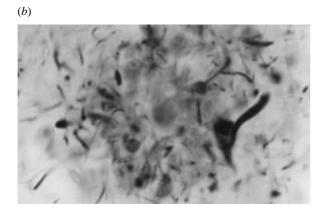


Figure 2. Light microscopy of (a) a neurofibrillary tangle in a pyramidal cell (note that the tangle fills the cell body as well as extending into the apical dendrite and the basal dendrites); (b) a senile plaque with a pale central core of amyloid surrounded by darkly stained swollen neurites.

(d) Neuropil threads

Neuropil threads are thread-like structures in the neuropil of the grey matter (Braak et al. 1986). Ultrastructurally and immunocytochemically, they have been shown to be essentially the same as the PHF found in neurofibrillary tangles and in the dystrophic neurites of neuritic plaques (Braak et al. 1986, 1994). They are therefore another aspect of neurofibrillary degeneration and in normal ageing are restricted mainly to the entorhinal cortex, hippocampus and amygdala, but are more widespread in Alzheimer's disease.

(e) Granulovacuolar degeneration

As the description implies, the lesion is of one or more apparently empty vacuoles other than for a central granule, which are found in the cell bodies usually of pyramidal cells in the hippocampus. The extent of this lesion increases with age and is more extensive in Alzheimer's disease (Xu et al. 1992). Little is known about the biochemical contents of granulovacuoles other than that the granules react with antibodies to cytoskeletal proteins, i.e. neurofilaments, tubulin and tau and also to ubiquitin (Kahn et al. 1985; Mayer et al. 1989; Dickson et al. 1993).

(f) Hirano bodies

Hirano bodies are rod-shaped structures up to 30 µm long and 8 µm wide that are found in or adjacent to

hippocampal pyramidal cells. They increase in number with age and are also more abundant in Alzheimer's disease. Ultrastructurally they appear as paracrystalline arrays of 60-100 nm filaments. Like PHF/straight filaments and granulovacuolar degeneration, Hirano bodies seem to be composed of cytoskeletal proteins but principally those proteins associated with microfilaments, i.e. tropomyosin, α actinin and vinculin (Galloway et al. 1987a), although antibodies to tau and neurofilament proteins have also been reported to stain Hirano bodies (Galloway et al. 1987b; Schmidt et al. 1989).

(g) Congophilic angiopathy or cerebral amyloid angiopathy

This is a change in which there is extracellular deposition of $A\beta$ in the walls of cerebral blood vessels. It is observed with increasing frequency with age and is extensive in Alzheimer's disease. The walls of severely affected blood vessels are thickened and associated with haemorrhage and other vessel wall damage. In rare families with a familial form, massive accumulation of $A\beta$ occurs, resulting in fatal haemorrhage (Wattendorff et al. 1995). However, the age-related changes in vessel walls are not just restricted to $A\beta$ deposition since there are also changes in the composition of connective tissue and smooth muscle as well as thickening of the vascular basement membrane (Kalaria 1996).

(h) Infarcts and leucoaraiosis

The accumulation of numerous small infarcts is possibly the second most common cause of dementia after Alzheimer's disease. Infarcts in small numbers are also found in brains from normal elderly individuals. Leucoaraiosis is a term used to describe a rarefaction of white matter usually close to the ventricles and can be visualized by computed tomography (CT) scanning (Nencini et al. 1993). The change tends to be diffuse and is more common in elderly brains and is present in Alzheimer's disease. The cause of the white matter changes are not known but may be the result of partial ischaemia.

3. DEMENTIA-NORMAL OR PATHOLOGICAL AGEING?

The brain, like other organs, is subject to increased incidence of disease as we age; however, it is dementia that is the most common form of pathological ageing and since the severity of dementia is correlated with increased levels of the changes described in the preceding section of this review, the remainder of this article will focus on current knowledge of the biological substrate of dementia, and particularly Alzheimer's disease. It is because the pathological changes of Alzheimer's disease are present in small numbers in the brains of intellectually intact individuals that the question has arisen of whether Alzheimer's disease is simply the inevitable outcome of normal ageing or if it is a disease. There is, as yet, no straightforward answer

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and, indeed, increasingly the question is probably oversimplistic in the light of newly discovered genetic risk factors.

Early onset Alzheimer's disease is usually an autosomal dominant condition and, so far, the identified loci are mutations in the gene for the precursor protein from which A β is derived, the amyloid precursor protein (APP), and in the genes for two related proteins, presenilin 1 (PS1) and presenilin 2 (PS2) (table 1). Mutations in APP account for perhaps 5% of familial Alzheimer's disease and mutations in *PS1* for 60–70%; *PS2* mutations are rare and so there is likely to be at least one more gene to be identified, mutations in which would cause autosomal dominant Alzheimer's disease.

Most Alzheimer's disease is late onset but it is now clear that much late onset Alzheimer's disease is associated with possession of the apolipoprotein ϵ 4 allelic form of the apolipoprotein (Roses 1996). There are three common alleles of apolipoprotein E, ϵ 2, ϵ 3 and ϵ 4, with ϵ 3 being the most frequent followed by ϵ 4 and, lastly, ϵ 2, which is relatively rare. Numerous studies of different ethnic groups have shown that the ϵ 4 allele is 2.5 times more frequent in Alzheimer's patients than in the general population and possession of two ϵ 4 alleles results in an earlier age of onset than one ϵ 4 allele, whereas, no ϵ 4 alleles is associated with the latest onset. However, apolipoprotein ϵ 4 is not essential for development of Alzheimer's disease since cases homozygous for apolipoprotein ϵ 3 are not uncommon.

It has also been reported that a polymorphism in the PS1 gene is associated with increased risk of lateonset Alzheimer's disease (Wragg et al. 1996; Kehoe et al. 1996), and it is highly likely that other genetic factors will be found to contribute risk. The risk of developing Alzheimer's disease is therefore probably determined by an interaction of the individual's genetic makeup with environmental factors, one example being head injury, which seems to be a risk factor (Schofield et al. 1997). Another putative environmental risk factor is herpes simplex virus since a recent study has found that possession of an apolipoprotein £4 allele only increases risk of Alzheimer's disease in those individuals with postmortem evidence of having had a latent herpes infection in the brain (Itzhaki et al. 1997); it is important that other laboratories confirm this observation because it would clearly have implications for understanding molecular mechanisms and possibly for therapy. Healthy or pathological ageing, more generally, is likely to be determined by this interplay between genotype and environment, as indeed will, perhaps, many if not most diseases, and within the next decade it should be possible by association studies to identify which genes and their alleles contribute risk to disease and confer predisposition for healthy ageing.

4. THE PATHOGENESIS OF SENILE PLAQUES AND NEUROFIBRILLARY TANGLES

Naturally, research effort into the molecular and cell biology of these lesions has concentrated on their devel-

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Table 1. Genetic factors in Alzheimer's disease and their biological consequences

type of genetic effect	biological consequences	
(a) autosomal dominant mutations	ì	
APP ₇₁₇ —three mutations of	increase in the ratio of	
valine to isoleucine,	$A\beta_{42}:A\beta_{40}$	
glycine or phenylalanine		
APP _{670,671} double mutation	five to eight-fold increase in	
	total Aβ	
presenilin-1 mutations	some demonstrated to	
	increase the ratio of	
	$A\beta_{42}$: $A\beta_{40}$	
presenilin-2 mutations	some demonstrated to	
	increase the ratio of	
	$A\beta_{42}$: $A\beta_{40}$	
one or more genes yet to be	5	
identified		
(b) genetic risk factors		
apolipoprotein ɛ4	promotes Aβ aggregation?	
	Fails to protect against tau	
	hyperphosphorylation?	
presenilin-1 polymorphism	unknown	
other genetic risk factors are	5	
likely to be discovered		

opment in Alzheimer's disease but since plaques and tangles in normal brain are indistinguishable from those in Alzheimer brain, the mechanisms underlying their formation are likely to be similar. These same lesions are also present in the brains of those affected by Down's syndrome but they develop at an earlier age, such individuals having abundant plaques and tangles in their fourth decade. However, examination of the brains from younger Down's syndrome cases has shown that diffuse $A\beta$ deposits occur in the late teens but neuritic plaques and tangles do not appear until 20 years later (Mann & Esiri 1989). This observation is a powerful argument for $A\beta$ deposition being an early event in pathogenesis and the presumed mechanism is that there is over-expression of the APP gene since it is located on chromosome 21, these individuals therefore having three copies of the gene. Thus, either over-expression of APP or expression of mutant forms of APP is a likely cause of A β production and deposition and recent experimental studies are consistent with such a mechanism, as discussed below.

(a) The amyloid precursor protein (APP) and production of $A\beta$;

APP is encoded by a single gene but alternative splicing gives rise to at least eight isoforms, all, apart from one, being transmembrane proteins (Selkoe 1994, 1996) (figure 3). The most common are APP₆₉₅, APP₇₅₁ and APP₇₇₀, the latter two possessing one or two extra domains in the extracellular portion of the molecule. One of these domains present in both APP₇₁₇ and APP₇₇₀ is a protease inhibitor known as the Kunitz protein inhibitor (KPI) domain. Cell surface APP has the majority of the molecule as an extracellular domain with a short cytoplasmic tail. The A β segment is

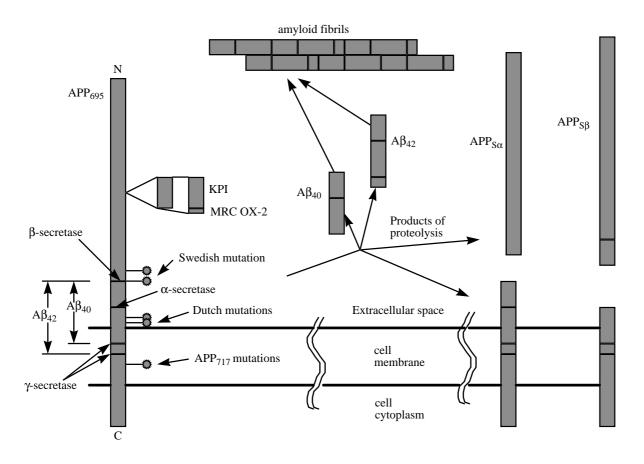


Figure 3. The structure and processing of APP. APP is a transmembrane protein shown here as APP_{695} ; APP_{751} includes the KPI domain and APP_{770} has both the KPI and MRC OX-2 domains. The locations of known mutations and secretase cleavage sites are shown, as are the main proteolytic products.

located at the interface of the extracellular domain and the membrane-spanning segment.

The normal function in the brain of APP is unknown although it is expressed by a variety of cells and in platelets the forms with KPI appear to play a role in haemostasis by inhibiting factor XIa (Smith et al. 1990). APP has, however, a rapid turnover and is proteolytically processed at several different sites (figure 3). One cleavage site is the so-called α -secretase site which is located within the $A\beta$ sequence. Cleavage at this site is referred to as non-amyloidogenic processing since full-length $A\beta$ cannot be generated by this pathway. Cleavage of APP at the N- and C-termini generates $A\beta$ and these sites are known as β and γ secretase sites. However, they are not unique sites since A β of different lengths can be produced, with species of 40 and 42 amino acids being the most abundant and/or important. Various studies have shown that minor species of $A\beta$ exist that are 'ragged' at both ends, giving rise to molecules that begin both N- and Cterminal of the main β -secretase site and, similarly, that end both N- and C-terminals of the main γ-secretase site.

Mutations in APP that give rise to autosomal dominant Alzheimer's disease flank the A β segment (Selkoe 1996). The first genetic lesions to be identified comprising three different point mutations are all located a few residues downstream of the γ -secretase site at position 717 of the APP₇₇₀ species (figure 3 and

table 1). A double mutation in a Swedish pedigree is in the two amino acids immediately N-terminal to the main β -secretase site. There are also a few Dutch and Flemish pedigrees in which mutations within the $A\beta$ segment at residues 692 and 693 give rise to massive amyloid angiopathy either without plaques and tangles or with these typical Alzheimer changes; these individuals frequently succumb to a fatal haemorrhage. It is assumed therefore that all of these mutations affect the proteolytic processing of APP at the three secretase sites, and indeed experimental studies using transfected cells has shown that the Swedish double mutation results in a 5-8 fold increase in secreted A β (Cai *et al.* 1993) and the APP₇₁₇ mutations result in an increase in the ratio of $A\beta_{42}$: $A\beta_{40}$ (Suzuki et al. 1994). Since $A\beta$ has a tendency to aggregate and because $A\beta_{42}$ has two additional hydrophobic amino acids compared to $A\beta_{40}$, then either a rise in total concentration of $A\beta$ or relatively more of the more hydrophobic $A\beta_{42}$ species can be predicted to drive aggregation.

Another major product of APP processing is the large extracellular domain, APP_S, that is secreted by cells. Depending upon whether this is derived by cleavage at the α - or β -secretase sites, at least two different species are produced (figure 3). APP_S has been found to have neuroprotective properties towards neurones exposed to a variety of insults, including excitotoxic levels of glutamate, and it has been reported that

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 $APP_{S\alpha}$ is more potent as a neuroprotectant than $APP_{S\beta}$ (Furukawa *et al.* 1996). This is a putative alternative mechanism to A\beta deposition and neurotoxicity (see below) as the initiating event in Alzheimer's pathogenesis in that changes in the levels of these two forms of APP_S could render neurones more vulnerable to other insults that might trigger subsequent changes in the brain resulting in Alzheimer's disease.

The subcellular sites in which APP is proteolysed are not established with certainty and may vary between cell types. Certainly, some APP is cleaved at the α secretase site at the cell surface but this is not the only location for this proteolytic event. Figure 4 illustrates schematically the current understanding of the trafficking and metabolism of APP from which it can be seen that some APP is processed to A β and APP_s in the endosomal compartment (Yamazaki *et al.* 1996; Koo *et al.* 1996).

Transgenic animal studies have also demonstrated the importance of A β for senile plaque formation. Most notably, mice transgenic for an *APP* minigene with one of the APP₇₁₇ mutations develop neuritic plaques remarkably similar to those typical of Alzheimer's disease (Games *et al.* 1995). However, these animals have so far not developed neurofibrillary tangles and so they appear to be a model for only part of the Alzheimer pathology (Masliah *et al.* 1996).

Experimental studies mainly using cultures of neurones have demonstrated that A β is neurotoxic but this property is dependent on the aggregation state of A β , fibrillar forms being more neurotoxic than amorphous A β (Busciglio *et al.* 1995; Yankner 1996; Mattson & Rydel 1996; Fraser *et al.* 1997; Iversen *et al.* 1995; Davis 1996). Since A β accumulates as diffuse deposits in the brains of young Down's syndrome patients and also in the non-demented elderly, there is still much to be learned about the properties of different forms of A β since in these examples of A β deposition in human brain, there is not likely to be significant neurotoxicity since symptoms of dementia are absent.

(b) The presenilins

The presenilins are serpentine membrane proteins traversing the membrane eight times (figure 5). They are located intracellularly mainly in the endoplasmic reticulum and Golgi apparatus and are proteolytically processed into at least two fragments, cleavage occurring in the large loop between two of the membranespanning segments (Doan *et al.* 1996; De Strooper *et al.* 1997). In excess of 35 mutations have been described in PSI in over 50 familial Alzheimer pedigrees; by contrast only two mutations have been found in PS2 (Hardy 1996).

Presenilin function is unknown but presenilins are homologous to two proteins, Sel-12 and Spe-4, in *C. elegans* (Sherrington *et al.* 1995; Levitan & Greenwald 1995; Levitan *et al.* 1996). The greatest homology is with Sel-12, which has been demonstrated on the basis of genetic experiments to have a role in Notch signalling. Notch is important in cell fate determination in development but must have a role in the adult since it is expressed in adult tissues. Spe-4 is involved in

membrane trafficking in spermatids of C. elegans. These facts, so far, have not proved to offer much insight into how mutant presenilins trigger the pathogenesis of Alzheimer's disease. However, experimental studies have shown that they may influence APP metabolism because fibroblasts from individuals with presenilin mutations secrete relatively more $A\beta_{42}$ compared to $A\beta_{40}$, as do cells transfected with mutant presenilins, and plasma from Alzheimer's patients possessing mutant presenilins also has relatively more $A\beta_{42}$ as do the brains from transgenic mice expressing such mutant presenilins (Scheuner et al. 1996; Duff et al. 1996; Borchelt et al. 1996). One view concerning the mechanism by which presenilins affect APP metabolism is that they may regulate trafficking of APP through internal membraneous compartments and that this results in changes in the proportion of APP cleaved by different γ -secretases, there being some evidence for the existence of multiple γ -secretases that may be located in different endosomal compartments (Citron et al. 1996).

(c) Paired helical filaments (PHF)

The main structural protein in PHF is the microtubule-associated protein tau. PHF-tau is in a hyperphosphorylated state compared to control postmortem brain tau, with up to 19 phosphates per molecule in PHF-tau. Many of the sites that are phosphorylated in PHF-tau are serine or threonine followed by a proline in the tau sequence (Morishima-Kawashima et al. 1995). Our laboratory was one of two first to identify independently glycogen synthase kinase-3 (GSK-3) as a kinase capable of generating in vitro a phosphorylation state of tau similar to PHF-tau, as assessed by a marked decrease in electrophoretic mobility and reactivity with a panel of monoclonal antibodies specific for phosphorylated epitopes present in PHF-tau, but not found in normal postmortem brain tau (Mandelkow et al. 1992; Hanger et al. 1992). Subsequently, it was demonstrated that in transfected cells both forms of GSK-3 (α and β —separate gene products) generate a similar PHF-like phosphorylated state of tau, whereas mitogen-activated protein (MAP) kinase that phosphorylates tau in vitro to a state like PHF-tau do not seem capable of generating this phosphorylated state in cells (Lovestone et al. 1994; Latimer et al. 1995).

(d) Tau phosphorylation is a critical process not only in neurodegeneration but also in neurodevelopment and the role of GSK-3 β

We and our collaborators (Brion *et al.* 1993) were also among the first to demonstrate that in the foetal brain, a proportion of tau is in a hyperphosphorylated state similar to PHF-tau, as judged by immunoreactivity with a panel of phosphorylation-sensitive monoclonal antibodies (Watanabe *et al.* 1993; Kenessey & Yen 1993; Hasegawa *et al.* 1993). Subsequently, it was shown that adult brain tau, when rapidly isolated, also contains a fraction of tau that is in a similarly hyperphosphorylated state (Matsuo *et al.* 1994). However, based upon

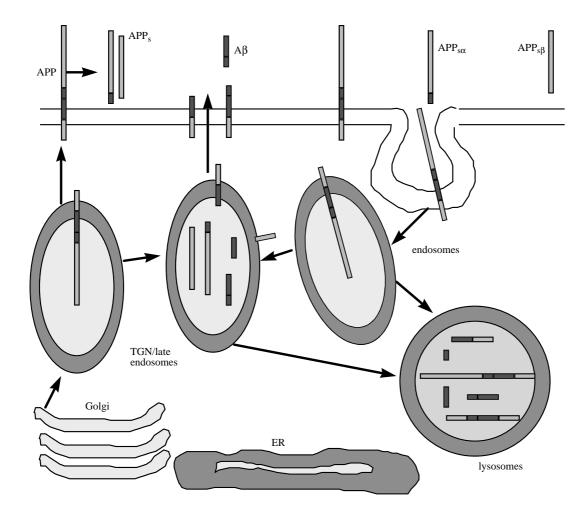


Figure 4. The trafficking and sites of processing of APP. APP is synthesized in the endoplasmic reticulum (ER) and passes through the Golgi apparatus to the trans-Golgi/late endosome compartments. Some APP is processed at the cell surface and other molecules are reinternalized after which they may be recycled to the cell surface or proteolysed and the products secreted; other APP molecules are probably completely degraded in the lysosomes.

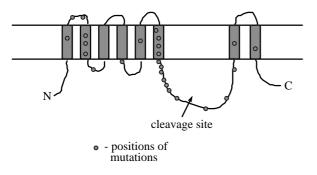


Figure 5. Presenilin structure. Presenilins are mainly located in internal membranes and the polypeptide traverses the membrane eight times. The approximate locations of sites of some of the known mutations are shown.

two-dimensional gel analysis (Sergeant *et al.* 1995) as well as SDS-PAGE (SDS-polyacrylamide gel electrophoresis) (B.H.A., G. Gibb, D. Davis and D. P. Hanger, unpublished observations), PHF-tau appears to be more heavily phosphorylated than the hyperphosphorylated fractions of foetal and normal adult tau. Thus, PHF-tau may be phosphorylated at sites additional to those in non-pathological forms of

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tau, for which there is some evidence (Shiurba *et al.* 1996; Hasegawa *et al.* 1996), or PHF-tau may be 'frozen' in a hyperphosphorylated state. Evidence in favour of the latter is demonstrated by the rapid increase in hyperphosphorylation of cellular tau induced by phosphatase inhibitors (Davis *et al.* 1995; Arendt *et al.* 1995), suggesting that phosphate on tau is turning over rapidly and that a proportion of normal tau is always hyperphosphorylated. Thus, PHF-tau may somehow become 'frozen' in a hyperphosphorylated state with phosphatases no longer able to remove phosphate, possibly because of aggregation of the tau into PHF which might be triggered by generation of a truly abnormally phosphorylated site or by factors such as proteoglycans (Goedert *et al.* 1996).

It is likely that the developmental regulation of tau phosphorylation is important because of the profound effect this has on microtubule dynamics, and it has been shown that the phosphorylation of tau by GSK-3 both *in vitro* and in cells affects microtubule properties. *In vitro*, tau promotes nucleation of microtubules, resulting in a population of shorter but more abundant microtubules and phosphorylation by GSK-3 abolishes this property of tau, which mutagenesis studies have

shown resides in several phosphorylation sites (Utton *et al.* 1997). In transfected non-neuronal cells, tau bundles microtubules and induces neurite-like extensions (Kanai *et al.* 1992), but hyperphosphorylation of tau by GSK-3 in co-transfected COS cells abolished the microtubule-bundling properties of tau with a concomitant reduction in microtubule stability (Lovestone *et al.* 1996*b*). Thus, phosphorylation of tau by GSK-3 clearly results in regulation of its microtubule-modulating properties and leads to the conclusion that increased hyperphosphorylation or induction of a 'frozen' hyperphosphorylated state may be a factor in the observed loss of assembled microtubules in PHF-afflicted neurones, accounting for the cytoskeleton-PHF transition (figure 6).

(e) Apolipoprotein E

There have been a number of attempts to explain the genetic risk of possessing an apolipoprotein £4 allele but so far none have resulted in convincing mechanisms. Apolipoprotein E4 protein has been claimed to promote aggregation of $A\beta$ more strongly than apolipoprotein E3 (Strittmatter et al. 1993), but other studies have found the opposite effect and this may depend upon the source of apolipoprotein E used for the experiments (LaDu et al. 1994). Similarly, it has been proposed that apolipoprotein E3 binds tau more strongly than apolipoprotein E4 and may therefore protect tau from hyperphosphorylation (Fleming et al. 1996). This is a somewhat surprising proposal because apolipoproteins are normally not expected to gain access to the cytoplasmic compartment of cells where tau is present. However, there are studies that show the presence of apolipoprotein E in the cytoplasm and our laboratory has reported that tau may have a role in the selective retention of apolipoprotein E3 over E4 in the cytoplasm (Lovestone et al. 1996a). More studies are required to elucidate the mechanism by which the apolipoprotein allotype influences the pathogenesis of Alzheimer's disease and in view of the recently reported combined association of herpes simplex virus and apolipoprotein £4 alleles accounting for increased risk of Alzheimer's disease, it may be that apolipoprotein E influences neuronal susceptibility to herpes infection (Itzhaki et al. 1997).

(f) The amyloid cascade

The amyloid cascade hypothesis (figure 7) proposes that $A\beta$ deposition in the brain is the trigger for the rest of the pathogenesis of Alzheimer's disease, including PHF formation (Hardy & Higgins 1992). Although there is now good experimental evidence in support of altered APP metabolism leading to senile plaques, there is little evidence in support of this also giving rise to PHF. Two groups have claimed that treatment of cultured neurones with $A\beta$ leads to increased tau phosphorylation (Busciglio *et al.* 1995; Takashima *et al.* 1996), although work in our laboratory has failed to confirm such observations (Davis *et al.* 1995). It may be time, therefore, to look for alternative links between APP/A β and tau hyperphosphorylation/PHF formation.

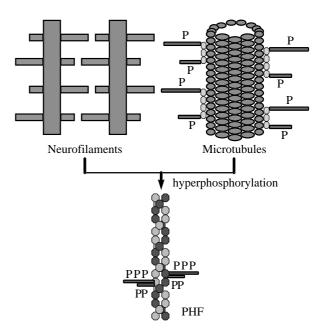


Figure 6. The cytoskeleton to PHF transition. The normal cytoskeleton in neurones consists principally of microtubules and neurofilaments. When PHF form, the cytoskeleton disappears and is replaced by PHF containing hyperphosphorylated tau.

Since phosphorylation of proteins is regulated by signal transduction events and it has been demonstrated by many laboratories that APP metabolism is modulated by activation of protein kinase C (PKC) (Efthimiopoulos et al. 1994; Nitsch et al. 1996; Govoni et al. 1996a), it may be timely to investigate the possible links between APP metabolism and tau phosphorylation that are mediated by signal transduction processes. There have been reports that PKC activity is depressed in Alzheimer brain and fibroblasts (Van-Huynh et al. 1989; Govoni et al. 1996a,b). A reduction in PKC activity would be compatible with increased amyloidogenic processing of APP and it has been reported that PKC activation reduces GSK-3β activity (Cook et al. 1996), thus again a reduction in PKC might lead to tau hyperphosphorylation. Future work may, therefore, be directed towards investigating if such a coupling between APP metabolism and tau phosphorylation via PKC occurs in neurones and in vivo.

5. OTHER RELATED NEURODEGENERATIVE DISEASES

Alzheimer's disease is the commonest neurodegenerative disease but other less common diseases may be related since several are characterized by neuronal inclusion bodies that are composed of cytoskeletal proteins, including tau (table 2). Since Alzheimer's disease possibly represents a form of accelerated or accentuated ageing, then these other conditions might also be similarly considered. There is evidence of Lewy bodies in the substantia nigra in putative presymptomatic Parkinson's disease (Gibb & Lees 1988).

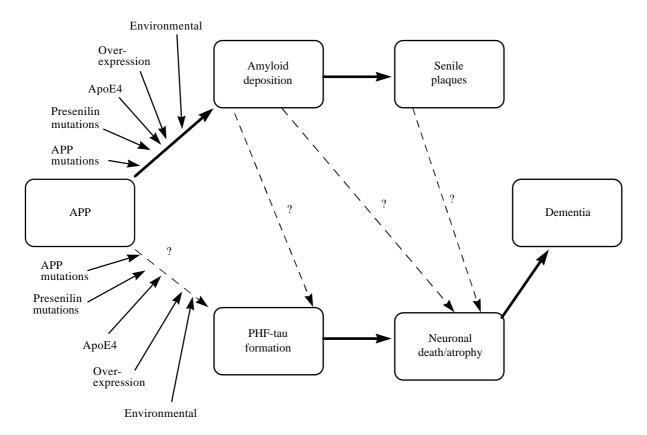


Figure 7. The amyloid cascade hypothesis. APP is presumed to be involved at an early stage and dementia is the result of neuronal loss or atrophy. Solid lines represent stages for which there is good experimental evidence whereas dotted lines indicate possible but unknown relationships.

Table 2. Neurodegenerative diseases-inclusion bodies and cytoskeletal constituents

neurodegenerative disease	inclusion body	ultrastructural component	main constituent cytoskeletal protein
Alzheimer's disease	neurofibrillary tangles	PHF and some straight filaments	tau
dementia with Lewy bodies	cortical Lewy bodies	neurofilaments	neurofilament triplet proteins
Parkinson's disease	Lewy bodies	neurofilaments	neurofilament triplet proteins
motor neurone disease	axonal spheroids and perikaryal inclusions	neurofilaments	neurofilament triplet proteins
Pick's disease	Pick bodies	straight filaments	tau
progressive supranuclear palsy (PSP)	PSP neurofibrillary tangles	straight filaments	tau
multiple system atrophy (MSA)	glial cytoplasmic inclusions	tubules	tau
corticobasal degeneration	neuronal inclusions	straight filaments	tau
Huntington's disease	none	-	

Dementia is also the consequence of neurodegeneration caused by some of these other rarer diseases such as multiple system atrophy or progressive supranuclear palsy, both of which have inclusions composed of tau (Jellinger 1996). Thus, with advancing age there may be an increasing risk of neurodegeneration and depending upon the interaction of genotype with environmental factors, one or other of these diseases of the nervous system may ensue.

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REFERENCES

- Arendt, T., Holzer, M., Fruth, R., Brückner, M. K. & Gärtner, U. 1995 Paired helical filament-like phosphorylation of tau, deposition of $\beta/A4$ -amyloid and memory impairment in rat induced by chronic inhibition of phosphatase 1 and 2A. Neuroscience 69, 691-698.
- Borchelt, D. R., Thinakaran, G., Eckman, C. B. & 16 coauthors. 1996 Familial Alzheimer's disease-linked presenilin 1 variants elevate A β 1-42/1-40 ratio in vitro and in vivo. Neuron 17, 1005-1013.
- Braak, H. & Braak, E. 1991 Neuropathological stageing of Alzheimer-related changes. Acta Neuropath. (Berl.) 82, 239-259.

BIOLOGICAL SCIENCES

- Braak, H., Braak, E., Grundke Iqbal, I. & Iqbal, K. 1986 Occurrence of neuropil threads in the senile human brain and in Alzheimer's disease: a third location of paired helical filaments outside of neurofibrillary tangles and neuritic plaques. *Neurosci. Lett.* 65, 351–355.
- Braak, E., Braak, H. & Mandelkow, E.-M. 1994 A sequence of cytoskeleton changes related to the formation of neurofibrillary tangles and neuropil threads. *Acta Neuropath. (Berl.)* 87, 554–567.
- Brion, J.-P., Smith, C., Couck, A.-M., Gallo, J.-M. & Anderton, B. H. 1993 Developmental changes in τ phosphorylation: Fetal τ is transiently phosphorylated in a manner similar to paired helical filament- τ characteristic of Alzheimer's disease. *J. Neurochem.* **61**, 2071–2080.
- Busciglio, J., Lorenzo, A., Yeh, J. & Yankner, B. A. 1995 βamyloid fibrils induce tau phosphorylation and loss of microtubule binding. *Neuron* 14, 879–888.
- Cai, X.-D., Golde, T. E. & Younkin, S. G. 1993 Release of excess amyloid β protein from a mutant amyloid β protein precursor. *Science* 259, 514–516.
- Chiu, H. C., Bondareff, W., Zarow, C. & Slager, U. 1984 Stability of neuronal number in human nucleus basalis of Meynert with age. *Neurobiol. Aging* 5, 83–88.
- Citron, M., Diehl, T. S., Gordon, G., Biere, A. L., Seubert, P. & Selkoe, D. J. 1996 Evidence that the 42- and 40-amino acid forms of amyloid β protein are generated from the βamyloid precursor protein by different protease activities. *Proc. Natn. Acad. Sci. USA* **93**, 13170–13175.
- Coleman, P. D. & Flood, D. G. 1987 Neuron numbers and dendrite extent in normal aging and Alzheimer's disease. *Neurobiol. Aging* 8, 521–545.
- Cook, D., Fry, M. J., Hughes, K., Sumathipala, R., Woodgett, J. R. & Dale, T. C. 1996 Wingless inactivates glycogen synthase kinase-3 via an intracellular signalling pathway which involves a protein kinase C. *EMBO J.* **15**, 4526–4536.
- Davis, D. R., Brion, J.-P., Couck, A.-M. & 8 co-authors. 1995 The phosphorylation state of the microtubule-associated protein tau as affected by glutamate, colchicine and βamyloid in primary rat cortical neuronal cultures. *Biochem. J.* **309**, 941–949.
- Davis, J. B. 1996 Oxidative mechanisms in β-amyloid cytotoxicity. *Neurodegen.* 5, 441–444.
- De Strooper, B., Beullens, M., Contreras, B. & 8 co-authors. 1997 Phosphorylation, subcellular localization, and membrane orientation of the Alzheimer's disease-associated presenilins. *J. Biol. Chem.* **272**, 3590–3598.
- Dickson, D. W. 1996 Senile cerebral amyloidosis (pathological aging) and cognitive status predictions: a neuropathology perspective. *Neurobiol. Aging* 17, 936–937.
- Dickson, D. W., Liu, W.-K., Kress, Y., Ku, J., DeJesus, O. & Yen, S.-H. C. 1993 Phosphorylated tau immunoreactivity of granulovacuolar bodies (GVB) of Alzheimer's disease: localization of two amino terminal tau epitopes in GVB. *Acta Neuropath. (Berl.)* 85, 463–470.
- Doan, A., Thinakaran, G., Borchelt, D. R. & 8 co-authors. 1996 Protein topology of presenilin 1. *Neuron* 17, 1023–1030.
- Duff, K., Eckman, C., Zehr, C. *et al.* 1996 Increased amyloid- β 42(43) in brains of mice expressing mutant presenilin 1. *Nature* **383**, 710–713.
- Efthimiopoulos, S., Felsenstein, K. M., Sambamurti, K., Robakis, N. K. & Refolo, L. M. 1994 Study of the phorbol ester effect on Alzheimer amyloid precursor processing: Sequence requirements and involvement of a cholera toxin sensitive protein. *J. Neurosci. Res.* 38, 81–90.
- Esiri, M. M., Hyman, B. T., Beyreuther, K. & Masters, C. L. 1997 Ageing and dementia. In *Greenfield's neuropathology* (ed. D. Graham & P. L. Lantos), pp. 153–233. Arnold.
- Flament-Durand, J. & Couck, A. 1979 Spongiform alterations in brain biopsies of presenile dementia. Acta Neuropath. (Berl.) 46, 159–162.
- Fleming, L. M., Weisgraber, K. H., Strittmatter, W. J., Troncoso, J. C. & Johnson, G. V. W. 1996 Differential

Phil. Trans. R. Soc. Lond. B (1997)

binding of apolipoprotein E isoforms to tau and other cytoskeletal proteins. *Exp. Neurol.* **138**, 252–260.

- Fraser, S. P., Suh, Y. H. & Djamgoz, M. B. A. 1997 Ionic effects of the Alzheimer's disease β -amyloid precursor protein and its metabolic fragments. *Trends Neurosci.* **20**, 67–72.
- Furukawa, K., Sopher, B. L., Rydel, R. E., Begley, J. G., Pham, D. G., Martin, G. M., Fox, M. & Mattson, M. P. 1996 Increased activity-regulating and neuroprotective efficacy of α-secretase-derived secreted amyloid precursor protein conferred by a C-terminal heparin-binding domain. *J. Neurochem.* 67, 1882–1896.
- Galloway, P. G., Perry, G. & Gambetti, P. 1987a Hirano body filaments contain actin and actin-associated proteins. *J. Neuropathol. Exp. Neurol.* 46, 185–199.
- Galloway, P. G., Perry, G., Kosik, K. S. & Gambetti, P. 1987b Hirano bodies contain tau protein. *Brain Res.* **403**, 337–340.
- Games, D., Adams, D., Alessandrini, R. & 21 co-authors. 1995 Alzheimer-type neuropathology in transgenic mice overexpressing V717F β-amyloid precursor protein. *Nature* 373, 523–527.
- Gibb, W. R. & Lees, A. J. 1988 The relevance of the Lewy body to the pathogenesis of idiopathic Parkinson's disease. *J. Neurol. Neurosurg. Psychiatry* **51**, 745–752.
- Goedert, M., Jakes, R., Spillantini, M. G., Hasegawa, M., Smith, M. J. & Crowther, R. A. 1996 Assembly of microtubule-associated protein tau into Alzheimer-like filaments induced by sulphated glycosaminoglycans. *Nature* 383, 550–553.
- Govoni, S., Bergamaschi, S., Gasparini, L. & 8 co-authors. 1996a Fibroblasts of patients affected by Down's syndrome oversecrete amyloid precursor protein and are hyporesponsive to protein kinase C stimulation. *Neurology* 47, 1069–1075.
- Govoni, S., Gasparini, L., Racchi, M. & Trabucchi, M. 1996b Peripheral cells as an investigational tool for Alzheimer's disease. *Life Sci.* 59, 461–468.
- Gray, E. G., Paula Barbosa, M. & Roher, A. 1987 Alzheimer's disease: paired helical filaments and cytomembranes. *Neuropathol. Appl. Neurobiol.* 13, 91–110.
- Hanger, D. P., Hughes, K., Woodgett, J. R., Brion, J.-P. & Anderton, B. H. 1992 Glycogen synthase kinase-3 induces Alzheimer's disease-like phosphorylation of tau: generation of paired helical filament epitopes and neuronal localisation of the kinase. *Neurosci. Lett.* 147, 58–62.
- Hansen, L. A., DeTeresa, R., Davies, P. & Terry, R. D. 1988 Neocortical morphometry, lesion counts, and choline acetyltransferase levels in the age spectrum of Alzheimer's disease. *Neurology* 38, 48–54.
- Hardy, J. 1996 New insights into the genetics of Alzheimer's disease. Ann. Med. 28, 255–258.
- Hardy, J. A. & Higgins, G. A. 1992 Alzheimer's disease: the amyloid cascade hypothesis. *Science* 256, 184–185.
- Harris, G. J., Schlaepfer, T. E., Peng, L. W., Lee, S., Federman, E. B. & Pearlson, G. D. 1994 Magnetic resonance imaging evaluation of the effects of ageing on greywhite ratio in the human brain. *Neuropathol. Appl. Neurobiol.* 20, 290–293.
- Hasegawa, M., Jakes, R., Crowther, R. A., Lee, V. M. Y., Ihara, Y. & Goedert, M. 1996 Characterization of mAb AP422, a novel phosphorylation-dependent monoclonal antibody against tan protein. *FEBS Lett.* **384**, 25–30.
- Hasegawa, M., Watanabe, A., Takio, K., Suzuki, M., Arai, T., Titani, K. & Ihara, Y. 1993 Characterization of two distinct monoclonal antibodies to paired helical filaments: further evidence for fetal-type phosphorylation of the τ in paired helical filaments. *J. Neurochem.* **60**, 2068–2077.
- Itzhaki, R. F., Lin, W. R., Shang, D. H., Wilcock, G. K., Faragher, B. & Jamieson, G. A. 1997 Herpes simplex virus type 1 in brain and risk of Alzheimer's disease. *Lancet* **349**, 241–244.

PHILOSOPHICAL TRANSACTIONS

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The ageing brain B. H. Anderton 1791

- Iversen, L. L., Mortishire-Smith, R. J., Pollack, S. J. & Shearman, M. S. 1995 The toxicity in vitro of β -amyloid protein. Biochem. J. **311**, 1–16.
- Jellinger, K. A. 1996 Structural basis of dementia in neurodegenerative disorders. J. Neural Transm. 103, 1–29.
- Kahn, J., Anderton, B. H., Probst, A., Ulrich, J. & Esiri, M. M. 1985 Immunohistological study of granulovacuolar degeneration using monoclonal antibodies to neurofilaments. *J. Neurol. Neurosurg. Psychiatry* 48, 924–926.
- Kalaria, R. N. 1996 Cerebral vessels in ageing and Alzheimer's disease. *Pharmac. Ther.* **72** 193–214.
- Kanai, Y., Chen, J. & Hirokawa, N. 1992 Microtubule bundling by tau proteins *in vivo*: analysis of functional domains. *EMBO J.* 11, 3953–3961.
- Kehoe, P., Williams, J., Lovestone, S., Wilcock, G. & Owen, M. J. 1996 Presenilin-1 polymorphism and Alzheimer's disease. *Lancet* 347, 1185.
- Kenessey, A. & Yen, S.-H. C. 1993 The extent of phosphorylation of fetal tau is comparable to that of PHF-tau from Alzheimer paired helical filaments. *Brain Res.* 629, 40–46.
- Koo, E. H., Squazzo, S. L., Selkoe, D. J. & Koo, C. H. 1996 Trafficking of cell-surface amyloid β-protein precursor. 1. Secretion, endocytosis and recycling as detected by labeled monoclonal antibody. *J. Cell Sci.* 109, 991–998.
- LaDu, M. J., Falduto, M. T., Manelli, A. M., Reardon, C. A., Getz, G. S. & Frail, D. E. 1994 Isoform-specific binding of apolipoprotein E to β -amyloid. *J. Biol. Chem.* **269**, 23403–23406.
- Latimer, D. A., Gallo, J.-M., Lovestone, S., Miller, C. C. J., Reynolds, C. H., Marquardt, B., Stabel, S., Woodgett, J. R. & Anderton, B. H. 1995 Stimulation of MAP kinase by vraf transformation of fibroblasts fails to induce hyperphosphorylation of transfected tau. FEBS Lett. 365, 42–46.
- Levitan, D., Doyle, T. G., Brousseau, D., Lee, M. K., Thinakaran, G., Slunt, H. H., Sisodia, S. S. & Greenwald, I. 1996 Assessment of normal and mutant human presenilin function in *Caenorhabditis elegans. Proc. Natn. Acad. Sci. USA* 93, 14940–14944.
- Levitan, D. & Greenwald, I. 1995 Facilitation of *lin-12*mediated signalling by *sel-12*, a *Caenorhabditis elegans* S182 Alzheimer's disease gene. *Nature* 377, 351–354.
- Lippa, C. F., Hamos, J. E., Pulaski Salo, D., Degennaro, L. J. & Drachman, D. A. 1992 Alzheimer's disease and aging: effects on perforant pathway perikarya and synapses. *Neurobiol. Aging* 13, 405–411.
- Lovestone, S., Reynolds, C. H., Latimer, D. & 9 co-authors. 1994 Alzheimer's disease-like phosphorylation of the microtubule-associated protein tau by glycogen synthase kinase-3 in transfected mammalian cells. *Curr. Biol.* 4, 1077–1086.
- Lovestone, S., Anderton, B. H., Hartley, C., Jensen, T. G. & Jorgensen, A. L. 1996a The intracellular fate of apolipoprotein E is tau dependent and apoe allele-specific. *Neuroreport* 7, 1005–1008.
- Lovestone, S., Hartley, C. L., Pearce, J. & Anderton, B. H. 1996b Phosphorylation of tau by glycogen synthase kinase-3beta in intact mammalian cells: the effects on the organization and stability of microtubules. *Neuroscience* 73, 1145– 1157.
- Mandelkow, E.-M., Drewes, G., Biernat, J., Gustke, N., van Lint, J., Vandenheede, J. R. & Mandelkow, E. 1992 Glycogen synthase kinase-3 and the Alzheimer-like state of microtubule-associated protein tau. *FEBS Lett.* **314**, 315–321.
- Mann, D. M. & Esiri, M. M. 1989 The pattern of acquisition of plaques and tangles in the brains of patients under 50 years of age with Down's syndrome. *J. Neurol. Sci.* 89, 169– 179.
- Mann, D. M. A. & Yates, P. O. 1979 The effects of ageing on the pigmented nerve cells of the human locus ceruleus and substantia nigra. *Acta Neuropath. (Berl.)* **47**, 93–97.

Phil. Trans. R. Soc. Lond. B (1997)

- Masliah, E., Mallory, M., Hansen, L., DeTeresa, R. & Terry, R. D. 1993 Quantitative synaptic alterations in the human neocortex during normal aging. *Neurology* **43**, 192–197.
- Masliah, E., Sisk, A., Mallory, M., Mucke, L., Schenk, D. & Games, D. 1996 Comparison of neurodegenerative pathology in transgenic mice overexpressing V717F βamyloid precursor protein and Alzheimer's disease. J. Neurosci. 16, 5795–5811.
- Matsuo, E. S., Shin, R.-W., Billingsley, M. L., Van DeVoorde, A., O'Connor, M., Trojanowski, J. Q. & Lee, V. M.-Y. 1994 Biopsy-derived adult human brain tau is phosphorylated at many of the same sites as Alzheimer's disease paired helical filament tau. *Neuron* 13, 989–1002.
- Mattson, M. P. & Rydel, R. E. 1996 Alzheimer's disease amyloid ox-tox transducers. *Nature* 382, 674–675.
- Mayer, R. J., Lowe, J., Lennox, G., Doherty, F. & Landon, M. 1989 Intermediate filaments and ubiquitin: a new thread in the understanding of chronic neurodegenerative diseases. *Prog. Clin. Biol. Res.* **317**, 809–818.
- Monagle, R. D. & Brody, H. 1974 The effects of age upon the main nucleus of the inferior olive in the human. *J. Comp. Neurol.* **155**, 61–66.
- Morishima-Kawashima, M., Hasegawa, M., Takio, K., Suzuki, M., Yoshida, H., Titani, K. & Ihara, Y. 1995 Proline-directed and non-proline-directed phosphorylation of PHF-tau. *J. Biol. Chem.* 270, 823–829.
- Nencini, P., Inzitari, D., Gibbs, J. & Mangiafico, S. 1993 Dementia with leucoaraiosis and dural arteriovenous malformation: clinical and PET case study. *J. Neurol. Neurosurg. Psychiatry* 56, 929–931.
- Nitsch, R. M., Deng, M. H., Growdon, J. H. & Wurtman, R. J. 1996 Serotonin 5-HT2a and 5-HT2e receptors stimulate amyloid precursor protein ectodomain secretion. *J. Biol. Chem.* 271, 4188–4194.
- Roses, A. D. 1996 Apolipoprotein E alleles as risk factors in Alzheimer's disease. A. Rev. Med. 47, 387–400.
- Scheuner, D., Eckman, C., Jensen, M. & 18 co-authors. 1996 Secreted amyloid β -protein similar to that in the senile plaques of Alzheimer's disease is increased *in vivo* by the presenilin 1 and 2 and *APP* mutations linked to familial Alzheimer's disease. *Nature Med.* **2**, 864–870.
- Schmidt, M. L., Lee, V. M. & Trojanowski, J. Q. 1989 Analysis of epitopes shared by Hirano bodies and neurofilament proteins in normal and Alzheimer's disease hippocampus. *Lab. Invest.* **60**, 513–522.
- Schofield, P.W., Tang, M., Marder, K., Bell, K., Dooneief, G., Chun, M., Sano, M., Stern, Y. & Mayeux, R. 1997 Alzheimer's disease after remote head injury: an incidence study. *J. Neurol. Neurosurg. Psychiatry* **62**, 119–124.
- Selkoe, D. J. 1994 Amyloid β-protein precursor: new clues to the genesis of Alzheimer's disease. *Curr. Opin. Neurobiol.* 4, 708–716.
- Selkoe, D. J. 1996 Amyloid β-protein and the genetics of Alzheimer's disease. *J. Biol. Chem.* **271**, 18295–18298.
- Sergeant, N., Bussière, T., Vermersch, P., Lejeune, J. P. & Delacourte, A. 1995 Isoelectric point differentiates PHFtau from biopsy-derived human brain tau proteins. *Neuroreport* 6, 2217–2220.
- Sherrington, R., Rogaev, E. I., Liang, Y. & 21 co-authors. 1995 Cloning of a gene bearing missense mutations in early-onset familial Alzheimer's disease. *Nature* **375**, 754–760.
- Shiurba, R. A., Ishiguro, K., Takahashi, M. & 8 co-authors. 1996 Immunocytochemistry of tau phosphoserine 413 and tau protein kinase I in Alzheimer pathology. *Brain Res.* 737, 119–132.
- Smith, R. P., Higuchi, D. A. & Broze, G. J. J. 1990 Platelet coagulation factor XIa-inhibitor, a form of Alzheimer amyloid precursor protein. *Science* 248, 1126–1128.
- Strittmatter, W. J., Weisgraber, K. H., Huang, D. Y., Dong, L.-M., Salvesen, G. S., Pericak-Vance, M., Schmechel, D., Saunders, A. M., Goldgaber, D. & Roses, A. D. 1993

Binding of human apolipoprotein E to synthetic amyloid β peptide: isoform-specific effects and implications for lateonset Alzheimer disease. *Proc. Natn. Acad. Sci. USA* **90**, 8098–8102.

- Suzuki, N., Cheung, T. T., Cai, X.-D., Odaka, A., Otvos, L. Jr, Eckman, C., Golde, T. E. & Younkin, S. G. 1994 An increased percentage of long amyloid β protein secreted by familial amyloid β protein precursor (βAPP717) mutants. *Science* **264**, 1336–1340.
- Takashima, A., Noguchi, K., Michel, G., Mercken, M., Hoshi, M., Ishiguro, K. & Imahori, K. 1996 Exposure of rat hippocampal neurones to amyloid β peptide (2535) induces the inactivation of phosphatidyl inositol-3 kinase and the activation of tau protein kinase I glycogen synthase kinase-3 β . *Neurosci. Lett.* **203**, 33–36.
- Terry, R. D., DeTeresa, R. & Hansen, L. A. 1987 Neocortical cell counts in normal human adult. Ann. Neurol. 21, 530– 539.
- Utton, M. A., Vandecandelaere, A., Wagner, U., Reynolds, C. H., Gibb, G. G., Miller, C. C. J., Bayley, P. M. & Anderton, B. H. 1997 Phosphorylation of tau by glycogen synthase kinase-3beta affects the ability of tau to promote microtubule self-assembly. *Biochem. J.* 323, 741–747.
- Van-Huynh, T., Cole, G., Katzman, R., Huang, K. P. & Saitoh, T. 1989 Reduced protein kinase C immunoreactivity and altered protein phosphorylation in Alzheimer's disease fibroblasts. *Arch. Neurol.* 46, 1195–1199.

- Watanabe, A., Hasegawa, M., Suzuki, M., Takio, K., Maroshima-Kawashima, M., Titani, K., Arai, T., Kosik, K. S. & Ihara, Y. 1993 *In vivo* phosphorylation sites in fetal and adult rat tau. *J. Biol. Chem.* 268, 25712–25717.
- Wattendorff, A. R., Frangione, B., Luyendijk, W. & Bots, G. T. A. M. 1995 Hereditary cerebral haemorrhage with amyloidosis, Dutch type (HCHWA-D): clinicopathological studies. *J. Neurol. Neurosurg. Psychiatry* 58, 699–705.
- West, M. J. 1993 Regionally specific loss of neurones in the aging human hippocampus. *Neurobiol. Aging* 14, 287– 293.
- Wragg, M., Hutton, M., Talbot, C. & 18 co-authors. 1996 Genetic association between intronic polymorphism in presenilin-1 gene and late-onset Alzheimer's disease. *Lancet* 347, 509–512.
- Xu, M., Shibayama, H., Kobayashi, H., Yamada, K., Ishihara, R., Zhao, P., Takeuchi, T., Yoshida, K., Inagaki, T. & Nokura, K. 1992 Granulovacuolar degeneration in the hippocampal cortex of aging and demented patients—a quantitative study. *Acta Neuropathol. (Berl.)* 85, 19.
- Yamazaki, T., Koo, E. H. & Selkoe, D. J. 1996 Trafficking of cell-surface amyloid β-protein precursor. 2. Endocytosis, recycling, and lysosomal targeting detected by immunolocalization. *J. Cell Sci.* 109, 999–1008.
- Yankner, B. A. 1996 Mechanisms of neuronal degeneration in Alzheimer's disease. *Neuron* 16, 921–932.

BIOLOGICAL